Things we haven't covered that are of general interest

Two-way factorial ANOVA

Randomized Complete Block Designs (RCBD) Quick introduction to these topics Goal is that you know some names if you want to pursue any topic Stat 5710 (Intro to Expt. Design) covers factorial ANOVA and RCBD in great depth

Two-way factorial designs:

Treatments are classified by two different factors

My language: A treatment is a unique thing "done" to an experimental unit

Collard study: 4 methods to measure riboflavin in cooked collards

Experimental unit: 100 gm of collard leaves

Observational unit: 100 gm of collard leaves

Freatment	Sample size	Chemical Clarification	Mean
А	Small $(0.25 \text{ gm})$	No	$\mu_a$
В	Small $(0.25 \text{ gm})$	Yes	$\mu_b$
$\mathbf{C}$	Large $(1.00 \text{ gm})$	No	$\mu_c$
D	Large $(1.00 \text{ gm})$	Yes	$\mu_d$

Two-way factorial designs: cell means model

Cell-means model: Fitting a mean for each treatment, followed by linear contrasts We already know how to do this:

Fit an ANOVA model where each treatment has a different mean Know how to answer various relevant questions

- Q: Do all treatments have the same mean? Overall F test (3 df) comparing 4 different means model to equal means model
- Q: Which means differ from which others? Pairwise comparisons, almost certainly with Tukey adjustment for multiple comparisons
- Q: What is the difference between Small and Large, averaged over Clarification Linear contrast:  $(\mu_a + \mu_b)/2 (\mu_c + \mu_d)/2$
- Q: What is the difference between Yes and No, averaged over Sample size Linear contrast:  $(\mu_b + \mu_d)/2 (\mu_a + \mu_c)/2$
- Q: Is the effect of sample size (large small) the same for Yes and No? Linear contrast:  $(\mu_d - \mu_b) - (\mu_c - \mu_a)$

Collard analysis: cell means approach:

Cell means: a (sn) b (sy) c (ln) d (ly) Pooled sd:  $s_p = 2.57, 8 \text{ df}$ 42.6 25.1 37.0 23.7

Effect $a(sn) b(sy) c(ln) d(ly)$ Estimate se	p								
	0.047								
Sample size 0.5 0.5 -0.5 -0.5 3.48 1.48	0.047								
Clarify $0.5 - 0.5 - 0.5 - 0.5 - 15.45 - 1.48 <$	< 0.0001								
Interaction 1 -1 -1 1 -4.17 2.97	0.20								
Interpretation / conclusions:									
No evidence of an interaction between Sample size and Clarification									
Smaller sample size has 3.48 larger concentration									
Clarifying the sample reduces concentration by 15.45									
Factorial designs: Vocabulary									
Factor: Variable being manipulated									
2 factors: Sample size and Chemical Clarification									
Levels: each value of a factor									
Small or Large for Size, No or Yes for Clarification									
Two-way factorial: 2 factors									
$2 \ge 2$ factorial: 2 factors, each with 2 levels									
$3 \ge 5$ factorial: 2 factors, one with 3 levels, other with 5									
Four-way factorial: 4 factors, e.g., $2 \ge 2 \ge 3 \ge 5$ factorial									
Complete: treatments are all combinations of all levels of the factors									
Complete $2 \ge 2$ factorial will have 4 treatments (like the Collard study)									
Complete $3 \ge 5$ factorial will have 15 treatments									
Balanced: all cells have same sample size									
Focus on the treatments									
label them by the combination of levels, and									
organize in two rows and two columns									
Dot in the subscript represents averaging over that subscript									
$\mu_{.n} = (\mu_{sn} + \mu_{ln})/2$									
Chemical Clarification Marginal Mean									
Sample Size No Yes									

Sample Size	No	Yes	
Small	$\mu_{sn}$	$\mu_{sy}$	$\mu_{s.}$
Large	$\mu_{ln}$	$\mu_{ly}$	$\mu_{l.}$
Marginal Mean	$\mu_{.n}$	$\mu_{.y}$	

More vocabulary:

Cell mean: mean for one treatment

Marginal mean (or LSMEAN): mean for a level **averaged over all levels of other factors** Main effect: difference in marginal means

Simple effect: difference between cell means

Two-way factorial designs: factor-based analyses

Simpler way to get the same results

Avoids writing contrast coefficients

Based on the 2x2 organization of treatment means

Use 2 subscripts to identify treatments,

*i*: which level of sample size (s or l), j which level of clarification (n or y)

$$Y_{ijk} = \mu + \alpha_i + \beta_j + \alpha\beta_{ij} + \varepsilon_{ijk}$$

 $\alpha_i$  main effects for factor A:  $\alpha_s$  shared by both  $\mu_{sn}$  and  $\mu_{sy}$ 

 $\beta_j$  main effects for factor B:  $\beta_n$  shared by both  $\mu_{sn}$  and  $\mu_{ln}$ 

 $\alpha\beta_{ij}$  interaction effects: unique to each cell of the treatment structure

Get SS and tests of A (main effect of A), of B (main effect of B) and AB (interaction) Use type III SS

Answer useful questions even if data aren't balanced

Same results as using linear contrasts, but much simpler to write code

Follow up with contrasts or multiple comparisons between marginal means

e.g., between levels of A or levels of B

Collard example: factor-based analysis

Effect	р	LSMEANS		Difference	se
Sample size	0.047	S: 33.8	L: 30.4	3.48	1.48
Clarify	< 0.0001	N: 39.8	Y: 24.4	15.4	1.48
Interaction	0.20			-4.17	2.97

Same test results, same estimates, same standard errors as the cell means approach

Interpretation / usual practice:

Why consider main effects instead of simple effects?

When no interaction or interaction is small, definitely use main effects:

Main effects are more precise:

- marginal means are averages of cell means
- main effects are averages of simple effects

Main effect: Small - Large, se = 1.48.

Simple effect: Small - Large in No Clarify: se = 2.10

Main effect has double the sample size (6 obs get Small sample size, 3 get Small/No) Main effects give simpler summaries:

one estimate of effect of Small - Large, not two simple effects

Main are estimates of each simple effect

Usual practice: test interaction

If n.s., interpret main effects, both as averages and simple effects

If signif. interaction, think about whether average is interpretable

Don't interpret main effects as estimates of simple effects

My practice: always leave interactions in the model

Maintains connection to contrasts in cell means analysis

and pooled error sd,  $s_p$ , estimated only from within-cell variability

There are other opinions and practices

usually based on regression interpretations, not comparisons of means

Randomized complete block designs (RCBD):

Same idea as pairing, but with more than 2 treatments

Identify groups of similar experimental units (blocks)

Examples: Parts of a field, litters of pigs, age& sex groups

Want small variability within block, large differences between blocks

Randomly assign treatments within blocks

Want small blocks, so usually each trt only once per block

Better to have more small blocks, than fewer large blocks

Analysis of data from an RCBD:

Model:

 $Y_{ij} = \mu + \alpha_i + \beta_j + \varepsilon_{ij}$ 

i identifies blocks,  $\alpha_i$  are block main effects

j identifies treatments,  $\beta_j$  are treatment main effects

When 2 eu's per block, equivalent to pairing

Similar but not same as 2 way factorial ANOVA:

No interaction term.

Common to have each treatment only once in each block

When only one rep, can't separate error and interaction

Fundamental difference in interpretation

Don't really care about blocks

In model because that's how the study was designed

Blocks not randomly assigned;

Treatments are randomly assigned (within each block)

In a 2 way factorial, combinations of factors are randomly assigned General practice:

Block effects are always included in the model

Even when appear to be small

Don't test significance of block effects

You use blocks because you expect them to be different

Can quantify effectiveness of your blocks (see 5710)

Can guide design for your next study

Treatment design and experimental design:

Every study has two separate design components

Treatment design: What is done to an eu: the treatments and their structure

Experimental design: How treatments are randomly assigned to eu's

Most of the studies we've seen: treatments randomly assigned to eu's

Paired and block designs: treatments assigned within pairs or blocks

These are two different experimental designs.

Different ways of randomly assigning trts to eu's

Many experimental designs (see 5710)

Treatment design is the one-way, two-way, or something else structure of trts Again, many others (see 5710)

Why care about the difference Allow you to combine ideas, e.g., two-way factorial trt with blocks Guide constructing a model for **your** study Mixed models: Models with more than one "source" of variability or more than one e.u. e.u. = Experimental unit: "thing" randomly assigned to a treatment Everything this semester has focused on studies with one size of e.u. subsampling (multiple observation units per e.u.) dealt with by averaging Example: pens with multiple cows per pen ou = cowif treatments only assigned to pens, eu = penCan average cows within pen to get one obs per pen, so ou now = pen A mixed model explicitly has two sources of variability among pens and among cows Estimate both. Useful for study planning: how many pens? how many cows per pen? What if two treatment factors and two sizes of eu? feed amount randomly assigned to pen hormone stimulus randomly assigned to cow Called a split-plot study - modern analyses use mixed models Similar issues (different models) when each eu observed repeatedly 2nd course, Stat 5710, spends a lot of time on mixed models To estimate multiple sources of variation (variance components) To analyze data from split plot studies To analyze repeated measures data To analyze multi-site or multi-year studies

Overview of the entire semester:

Statistical analysis is a set of principles, not a set of recipes

- A good analysis is motivated by three things: your question(s) the study design, and the characteristics of the data
- Diagram from first lecture: Question → Data → Answers Design is how to collect data that answers your question Analysis is how to provide appropriate answers to your questions
- Goal is answers that are defensible because the statistical method is appropriate At least approximately
- Different models answer different questions Construct a model that answers **your** questions
- Estimates and confidence intervals are more useful answers than p-values
- Check your assumptions, but remember that real data only approximately matches assumptions
- Violating some assumptions doesn't make an analysis wrong Better fit to assumptions is almost always a better analysis
- Violating independence is usually trouble But random assignment of treatments can provide independence
- Almost everything else is details
- We have covered the core methods and principles There are lots of specialized areas of statistics each with their own sets of methods